

JOURNAL OF ANIMAL SCIENCE

The Premier Journal and Leading Source of New Knowledge and Perspective in Animal Science

Effects of B vitamin injections on plasma B vitamin concentrations of feed-restricted beef calves infected with bovine herpesvirus-1

P. L. Dubeski, F. N. Owens, W. O. Song, S. P. Coburn and J. D. Mahuren

J Anim Sci 1996. 74:1358-1366.

The online version of this article, along with updated information and services, is located on the World Wide Web at:

<http://jas.fass.org>



American Society of Animal Science

www.asas.org

Effects of B Vitamin Injections on Plasma B Vitamin Concentrations of Feed-Restricted Beef Calves Infected with Bovine Herpesvirus-1¹

P. L. Dubeski², F. N. Owens³, W. O. Song⁴, S. P. Coburn⁵, and J. D. Mahuren⁵

Department of Animal Science, Oklahoma State University, Stillwater 74078-0425

ABSTRACT: For nonruminants, stress and disease greatly increase requirements for vitamin B₆, folic acid, pantothenic acid, and ascorbate. The effects of feed restriction, virus infection, and vitamin injections on plasma concentrations of B vitamins critical to the immune response were evaluated. Twelve beef steer calves, 6 to 8 mo of age, were fed below maintenance for 17 d and deprived of food for 3 d during a 20-d period after weaning. They then were inoculated intranasally with live attenuated bovine herpesvirus-1 (BHV-1). Six calves received saline injections and six received injections of a B vitamin mixture and ascorbate every 48 h for 14 d before and 14 d after inoculation. A mild respiratory infection developed in

all calves 4 to 5 d after inoculation. In control calves, restricted intake and food deprivation decreased plasma vitamin B₆ and pantothenate and increased vitamin B₁₂ but did not affect folic acid and ascorbate concentrations. Vitamin injections increased plasma concentrations of vitamin B₆, folic acid, vitamin B₁₂, pantothenic acid, and ascorbate ($P < .002$). Plasma concentrations of vitamin B₆, vitamin B₁₂, pantothenic acid, and ascorbate, but not folic acid, were markedly reduced in all calves during the BHV-1 infection ($P = .001$). The vitamin B₆, pantothenic acid, vitamin B₁₂, and ascorbate status of stressed calves may affect their immune response to vaccination or infection.

Key Words: Immunity, Bovine Herpesvirus, Stress, Pyridoxal, Vitamins

J. Anim. Sci. 1996. 74:1358-1366

Introduction

Stress during transport and marketing can compromise the immune system and predisposes cattle to develop certain infectious diseases, including the bovine respiratory disease complex. Transport is an acute stressor for cattle and can elevate serum cortisol concentrations for 4 to 7 d (Von Tungeln, 1986). Cortisol and other glucocorticoids suppress the immune response in cattle and other species (Golub and Gershwin, 1985; Roth, 1985).

Feed and water intake often are decreased during marketing and transport of cattle, reducing the supply of energy and of nutrients synthesized by ruminal microbes, including the B vitamins. At the same time,

requirements for water-soluble vitamins critical to the immune response, particularly vitamin B₆, folic acid, pantothenic acid, and ascorbate, may be increased substantially by stress. In nonruminants, pantothenic acid and ascorbate are required for glucocorticoid synthesis (Goodman, 1960; Dvorak, 1984) and vitamin B₆ is important in regulation of glucocorticoid function (Allgood et al., 1990; Maksymowych, 1990). Mueller and Thomas (1975) estimated that stress or "moderate" injury may increase requirements of vitamin B₆ and folic acid by 8 to 15 times, respectively, and may double or quadruple requirements for vitamin B₁₂ and pantothenic acid, respectively. Supplementation with ascorbate and vitamin B₆ have helped to overcome cortisol-induced immunosuppression in both humans and experimental animals.

This study was designed to investigate short-term changes in plasma concentrations of several B vitamins that are critical to the immune response and that may be depleted during stress and disease. With other species, plasma vitamin concentrations are used routinely to assess B vitamin deficiencies and status (Jaffe, 1984; Sauberlich, 1984b). Although plasma B vitamin concentrations of ruminants have rarely been assessed, one might expect plasma changes to reflect alterations in vitamin status. Plasma concentrations of folic acid, vitamin B₁₂, pantothenic acid, vitamin

¹We thank Gene Komer of American Veterinary Products, Inc., Fort Collins, CO 80524 for providing custom formulations for B vitamins and ascorbic acid.

²Present address: Agriculture and Agri-Food Canada, Lacombe Research Centre, Lacombe, Alberta, Canada T4L 1W1.

³To whom correspondence should be addressed.

⁴Dept. of Food Sci. and Human Nutr., Michigan State Univ., East Lansing 48824.

⁵Biochem. Dept., Fort Wayne Hospital and Training Center, Fort Wayne, IN 46815.

Received July 21, 1995.

Accepted February 1, 1996.

B₆, and ascorbate were measured throughout the study to investigate the effects of vitamin injections, restricted feeding and food deprivation, and BHV-1 infection.

Materials and Methods

Animal Care. The experiment protocol was reviewed and approved by the Oklahoma State University Animal Care and Use Committee.

Experimental Procedure. Twelve Hereford × Angus calves 6 to 8 mo of age were used in the study. Each day of the study will be designated relative to the day that calves were inoculated with bovine herpesvirus-1. The study began when calves were weaned on April 23, 1991 (d -21). During the 20 d following weaning, the calves were adapted to their surroundings and to B vitamin injections. During the trial, each calf received prairie grass hay at 1% of its initial BW daily on an as-fed basis until d 2, except during a 3-d period of food deprivation (d -8 to d -6) to simulate conditions during marketing and transport. The feeding rate was increased to 1.5% of initial BW on d 3 until the end of the trial so that calves would maintain BW. The calves were inoculated intranasally with an attenuated vaccine strain of bovine herpesvirus-1 on d 0. Calves were stressed by weaning, the prolonged period of restricted feeding including 3 d of food deprivation, and infection with bovine herpesvirus-1 (BHV-1). Bovine herpesvirus-1 was used as a model for the bovine respiratory disease complex because it can cause a primary respiratory tract infection and predisposes cattle to *Pasteurella* infection. However, use of an attenuated vaccine strain of BHV-1 in this study was equivalent to a vaccination stress rather than acute disease stress. The detailed experimental procedures were described (Dubeski et al., 1996).

Vitamin Treatments. The calves were assigned randomly to two treatment groups (control, +Vit) with six calves per treatment. Each calf was injected i.m. with B vitamins and ascorbate (+Vit) or sterile saline (controls) every 48 h from d -11 to d 14 of the experiment. The B vitamin and ascorbate solutions were custom-formulated (American Veterinary Products, Fort Collins, CO). Vitamin concentrations, dosages given, and estimated requirements extrapolated from requirements of growing swine on a metabolic body weight basis are presented in Table 1.

Blood Samples. Blood samples (50 mL) were collected from each animal in 10-mL evacuated glass tubes (Vacutainer, Sherwood Medical, St. Louis, MO), 30 mL with potassium EDTA and 20 mL with sodium heparin as the anticoagulant. As presented in Table 2, blood samples were collected immediately before feeding at seven different times from each animal. Although vitamin injections were discontinued on d

Table 1. B vitamin requirements and amounts supplied by injection

Vitamin	Estimated requirement ^a	Stress factor ^b	Dosage ^c
Thiamin	6.76 mg	—	13.5 mg
Riboflavin	16.91 mg	—	33.8 mg
Niacin	67.64 mg	—	135.0 mg
Folic acid	2.09 mg	15	60.0 mg
Pantothenic acid	54.1 mg	2	216.0 mg
Vitamin B ₆	6.76 mg	8	108.0 mg
Vitamin B ₁₂	67.64 µg	2	270.0 µg
Sodium ascorbate	Unknown	—	1,000.0 mg

^aDaily B vitamin requirements for a 190.5-kg calf were estimated on a metabolic body weight basis as equivalent to 3.56 times the requirements for a 34-kg pig (NRC, 1988).

^bThe estimated daily requirement was multiplied by factors of 2 for pantothenic acid and vitamin B₁₂, 15 for folic acid, and 8 for vitamin B₆ to account for the increased requirements of these specific B vitamins during stress or "moderate" injury (Mueller and Thomas, 1975).

^cTwice the daily estimated requirement was supplied by injection every 2 d, with one 3-mL dose containing the B vitamins and one 4-mL dose containing sodium ascorbate. Controls received identical doses of sterile saline solution.

14, additional blood samples were collected on d 16 and d 18 from +Vit calves to monitor withdrawal effects. When blood samples for B vitamin analysis were taken on the same day that B vitamins were injected, blood samples were obtained < 1 h before vitamins were injected.

Plasma was separated from blood after centrifugation at 1,500 × g and was frozen (-20°C) in small aliquots for subsequent analysis of the vitamins. Samples were processed under subdued lighting to minimize photodegradation of the B₆ vitamins and ascorbate. For ascorbate analysis, heparinized plasma was mixed with fresh 10% (wt/vol) metaphosphoric acid (1:1) and frozen immediately.

Vitamin Analysis. Ascorbate in heparinized plasma was measured colorimetrically after derivitization with 2,4-dinitrophenylhydrazine (McCormick, 1986).

Table 2. Plasma sampling dates, number of vitamin injections, and time since last injection

Experimental day	Number of previous vitamin injections	Days since last vitamin injection
-45 (before weaning)	0	—
-15 (after weaning) ^a	0	—
-8 (feed deprivation began)	2	2
-5 (feed deprivation ended)	4	1
0 (BHV-1 inoculation)	6	2
5 (BHV-1 infection peak)	9	1
14 (final injection)	13	2
16 (after injections)	13	4
18 (after injections)	13	6

^aCalves were weaned on d -21.

Plasma concentrations of both folate and vitamin B₁₂ were measured in plasma (containing potassium EDTA) using a radioassay method (Quantaphase Folate and Vitamin B₁₂ Radioassay, Bio-Rad Clinical Division, Hercules, CA). Pantothenic acid in heparinized plasma was measured using a radioimmunoassay (Wyse et al., 1979). B₆ vitamins in heparinized plasma were analyzed using a cation-exchange HPLC procedure (Coburn and Mahuren, 1983).

Statistical Analysis. Plasma vitamin data were analyzed by ANOVA using the GLM procedure of SAS (1985) for the completely randomized design (all animals were inoculated but only half received vitamin injections) for the main effects of vitamin treatment. Because some of the samples were obtained before and some after inoculation, averages across collection dates would not be meaningful. Consequently, effects of vitamin injection were tested within each individual collection date. Changes from one date to another also were examined. In each case, the effect of vitamin injection was tested within each sample period using animal within treatment as the error term.

Results and Discussion

Animal Health and Stress

Details of animal feed intake, weight loss, immune measurements, and clinical signs are reported separately (Dubeski et al., 1996). The stress model used in this study (weaning, a long-term period of moderate feed restriction, 3 d of food deprivation after adaptation, no long-distance shipping) probably

resulted in less extreme physical and nutritional changes than occur in many shipping-stressed calves. Calves lost 7.7% of BW gradually during the first 20 d of the study. This is similar to the weight loss for many cattle during a single haul, but considerably less severe than a BW loss of 11% or more observed in highly stressed cattle (Griffin, 1983). Furthermore, these cattle were not stressed by transport, one of the most severe stressors for cattle, which elevates plasma cortisol for 4 to 7 d (Crookshank et al., 1979). Even after prolonged feed restriction, calves in this study had higher concentrations of plasma folic acid, vitamin B₁₂, pantothenic acid, vitamin B₆, and ascorbate than plasma concentrations of shipping-stressed calves in a previous survey (Dubeski, 1992). This might suggest that stress has a larger impact than restricted feed intake on plasma vitamin concentrations and perhaps on vitamin status.

Bovine herpesvirus-1 infection caused mild respiratory disease in all calves, apparently limited to the upper respiratory tract. Clinical symptoms peaked at 4 to 6 d after infection. The infection was equivalent to vaccination with a live attenuated virus strain, and the immune response was comparable to a successful vaccination except that humoral immunity (mean IgG titer to BHV-1 at 14-d after infection) seemed to be greater in vitamin-injected calves (1,120 vs 550, $P = .115$).

Folic Acid

Plasma concentrations are presented in Table 3. Plasma folate tended to be greater in calves after than before weaning (26.7 vs 15.4 nM). Plasma folate concentrations in stressed calves, feedlot steers, and lactating dairy cows averaged 22.0, 58.9, and 29.7 nM,

Table 3. Folic acid concentration in plasma (nM)

Day of experiment	Treatment mean ^a		SEM	Prob.
	Control	+Vit		
-45 Before weaning	15.4	15.4	3.8	—
-15 After weaning	26.7	26.7	2.9	.97
-8 Feed restriction began	29.4	64.1	5.4	.002
-5 Feed restriction ended	29.7	74.3	4.5	.001
0 BHV-1 inoculation	22.9	930.5	145.6	.002
5 Infection peak	24.7	66.6	4.8	.001
14 Final vitamin injection	27.6	63.9	5.9	.002
16 After end of injections	NA ^b	86.0	—	—
18 After end of injections	NA ^b	55.9	—	—
Contrast or change				
B vitamins (d -8 vs d -15)	5.6	37.4	5.2	.003
Feed deprivation (d -5 vs d -8)	-2.2	10.2	7.0	.32
Refeeding (d 0 vs d -5)	-6.8	856.2	147.2	.002
BHV-1 (d 5 vs d 0)	1.8	-864.1	149.0	.002
Recovery (d 14 vs d 5)	2.7	-2.7	5.0	.45

^an = 6.

^bNot analyzed.

respectively, in samples obtained from various classes of cattle (Dubeski, 1992). Plasma folic acid concentration in the control calves did not decrease during food deprivation.

B vitamin injections increased ($P = .02$) mean plasma folic acid concentration on d -8 and thereafter. The folic acid dose was equivalent to 60 mg/d, 15 times the dietary requirement (Table 1). Mean plasma folate in +Vit calves ranged from 55.9 to 86.0 nM (except pre-injection and on d 0), which is similar to the range for plasma folate concentrations of feedlot steers (32.2 to 79.7, mean 58.9 nM; Dubeski, 1992). Girard et al. (1989) injected 2-wk-old heifers with 2.5, 5.0, 10, and 20 of mg folic acid and successfully increased serum folate from an initial concentration of 18.6 to 33.5 nM, the amount found in 4-mo-old heifers. However, these relatively low dosages of folic acid did not increase serum folate concentrations in 4-mo-old heifers.

The extremely high plasma folic acid concentration (930.5 nM) in +Vit calves on d 0 is difficult to explain. These samples were taken 2 d after the previous B vitamin injection and immediately before BHV-1 inoculation. Extremely high concentrations of other vitamins (vitamin B₁₂, pantothenic acid, vitamin B₆, but not ascorbate) were observed on d 0. These samples were analyzed several times and at different dilutions in order to rule out experimental error.

Plasma folic acid was not decreased by the mild BHV-1 infection. Concentrations were slightly higher in controls on d 5 vs d 0, whereas in calves receiving vitamin injections, concentrations dropped drastically from d 0 to d 5 (interaction of vitamin injection with plasma concentration change of $P = .002$). If BHV-1 increases folic acid requirements, a decrease in plasma folic acid would be expected in the control calves. Folic

acid concentrations on d 5 for both treatment groups were very similar to their respective concentrations on d -8, d -5, and d 14. Relatively high plasma folic acid concentrations (14.5 to 33.1 nM) in shipping-stressed calves (Dubeski, 1992) also suggest that folic acid status, as appraised by plasma folate concentration, was not seriously decreased by stress and reduced feed intake.

Vitamin B₁₂

Plasma vitamin B₁₂ concentrations are presented in Table 4. Plasma concentrations of vitamin B₁₂ tended to be greater after than before weaning. Generally, values during the trial were higher than means from stressed calves, feedlot steers, and lactating dairy cows (185, 118, and 211 pM) from the survey conducted by Dubeski (1992).

Short-term changes in plasma B₁₂ have not been investigated except in response to injection. Plasma B₁₂ concentration in control calves increased during the period of injections (d -8 and after) and reached a maximum concentration on d 0. Plasma concentrations tended to be increased with food deprivation and during infection, probably due to release of vitamin B₁₂ from liver reserves. The liver contains high concentrations of vitamin B₁₂ (Saubertlich, 1990). Liver tissue is catabolized during food or water deprivation (Shorthose and Wythes, 1988).

An increased vitamin B₁₂ requirement during BHV-1 infection may have been responsible for the decreases in vitamin B₁₂ concentration between d 0 to d 5. Plasma B₁₂ dropped 31% in controls. The reasons for the decrease in plasma B₁₂ following BHV-1 administration are not known, but they could reflect the interrelationships between B₁₂ and folate, methionine and choline metabolism (Nauss and Newberne,

Table 4. Vitamin B₁₂ concentration in plasma (pM)

Day of experiment	Treatment mean ^a		SEM	Prob.
	Control	+Vit		
-45 Before weaning	169	169	26	
-15 After weaning	220	214	24	.86
-8 Feed restriction began	222	326	36	.07
-5 Feed restriction ended	275	526	36	.001
0 BHV-1 inoculation	301	1,597	149	.001
5 Peak BHV-1 infection	207	456	24	.001
14 Final vitamin injection	277	541	30	.001
16 After end of injections	NA ^b	491	—	—
18 After end of injections	NA ^b	418	—	—
Contrast or change				
B vitamins (d -8 vs d -15)	11	112	33	.06
Feed deprivation (d -5 vs d -8)	42	199	17	.001
Refeeding (d 0 vs d -5)	26	1,071	117	.001
BHV-1 (d 5 vs d 0)	-94	-1,141	142	.001
Recovery (d 14 vs d 5)	70	85	17	.54

^an = 6.

^bNot analyzed.

1981), and an increased B₁₂ requirement for cellular proliferation in the immune response. Plasma B₁₂ concentration had increased to 277 pM at d 14, which may reflect a decreased requirement, an increase in ruminal synthesis related to the higher feed intake on d 3 to d 14 (Zinn et al., 1987), or changes in liver storage or release of the vitamin.

The role of vitamin B₁₂ status in resistance to disease is not clear. Research in humans has centered on the autoimmune phenomena in pernicious anemia. Animal studies are limited because of difficulties in producing a B₁₂ deficiency in experimental animals. Because of the extensive ruminal production of B₁₂ and its analogs when cattle are fed Co-deficient diets and(or) high-concentrate diets, vitamin B₁₂ deficiencies may develop more quickly in ruminants than in humans or laboratory animals. Consequently, the ruminant could be a useful model for B₁₂ metabolism.

In +Vit calves, plasma B₁₂ tended to be greater than in controls at d -8 ($P = .07$) and was markedly elevated ($P < .01$) in +Vit calves on subsequent days. As occurred for plasma folate, B₁₂ concentrations were extremely high on d 0 (1,597 vs 301 pM in +Vit vs control calves, respectively). Plasma B₁₂ concentration tended to be slightly higher on d 14 than on d 5 but similar to the means on earlier days.

Pantothenic Acid

Plasma concentrations of pantothenic acid tended to be greater after than before weaning (Table 5). Plasma values in this study were greater than mean values for transport-stressed calves and lactating dairy cows (.095 and .089 μM) but more similar to those of feedlot steers (.143 μM) from the survey of Dubeski (1992). The low concentrations in stressed

calves may reflect the role of pantothenic acid in adrenal function and corticosteroid synthesis (Goodman, 1960; Fidanza et al., 1978). A stress model similar to that used in this experiment did not induce the high, sustained serum cortisol concentrations found in shipping-stressed calves (d'Offay and Rosenquist, 1988); the added stress of transport may be responsible for more extreme decreases in plasma pantothenate.

In contrast to folate and vitamin B₁₂, plasma pantothenic acid concentrations in control calves tended to be decreased during food deprivation. Plasma concentrations were consistently but not significantly greater in calves receiving vitamin injections.

Plasma pantothenate may be more sensitive to stress factors such as BHV-1 infection than to the supply of pantothenic acid. In unstressed rats, a dietary pantothenate deficiency may take 2 to 3 wk to affect plasma pantothenate concentrations (Song et al., 1990). However, in the current study, the mild BHV-1 infection markedly reduced plasma pantothenate concentrations within 5 d.

Bovine herpesvirus-1 infection resulted in lower plasma pantothenate concentrations on d 5 than at other times for both treatment groups ($P < .001$). In both groups, plasma pantothenate increased on d 14, possibly due partly to increased feed intake as well as recovery from BHV-1 infection.

Vitamin B₆

Due to the complexity of analysis, B₆ vitamers were measured only in plasma samples taken on d -15, 0, and 5 (Table 6). Total B₆ was similar in both treatment groups before B vitamin treatments began

Table 5. Pantothenic acid concentration in plasma (μM)

Day of experiment	Treatment mean ^a		SEM	Prob.
	Control	+Vit		
-45 Before weaning	.128	.128	.019	—
-15 After weaning	.196	.183	.017	.60
-8 Feed deprivation began	.137	.143	.015	.79
-5 Feed deprivation ended	.122	.156	.013	.11
0 BHV-1 inoculation	.159	.771	.079	.001
5 Peak BHV-1 infection	.100	.113	.037	.75
14 Final vitamin injection	.164	.179	.013	.42
16 After vitamin injection	NA ^b	.19	—	—
18 After vitamin injection	NA ^b	.20	—	—
Contrast or change				
B vitamins (d -8 vs d -15)	-.059	-.040	.021	.53
Feed deprivation (d -5 vs d -8)	-.014	.013	.014	.20
Refeeding (d 0 vs d -5)	.030	.611	.074	.001
BHV-1 (d 5 vs d 0)	-.029	-.676	.090	.001
Recovery (d 14 vs d -5)	.033	.066	.043	.60

^an = 6.

^bNot analyzed.

Table 6. B₆ vitamer concentration in plasma (nM)

Day of experiment	B ₆ vitamer	Treatment mean ^a		SEM	Prob.	
		Control	+Vit			
-15 (before vitamin treatments)	Pyridoxal phosphate (PLP)	65.3	-35.2	10.6	.07	
	Pyridoxal (PL)	124.7	133.2	20.4	.78	
	Pyridoxine (PN)	0.0	0.0	0.0	1.00	
	Total B ₆	190.0	168.4	20.6	.53	
	4-Pyridoxic acid (4-PA)	25.0	24.8	2.2	.96	
0 (BHV-1 Inoculation)	Pyridoxal phosphate	56.8	55.7	10.7	.94	
	Pyridoxal	93.2	645.2	42.4	.001	
	Pyridoxine	0.0	1,822.3	305.4	.002	
	Total B ₆	150.0	2,523.2	180.2	.001	
	4-Pyridoxic acid	32.7	2,060.0	133.5	.001	
5 (Peak BHV-1)	Pyridoxal phosphate	68.8	91.5	22.2	.49	
	Pyridoxal	63.0	95.0	4.9	.001	
	Pyridoxine	0.0	9.7	6.8	.34	
	Total B ₆	131.8	196.2	21.5	.04	
	4-Pyridoxic acid	23.2	51.2	10.7	.10	
Contrasts and interactions		PLP	PL	PN	B ₆	4-PA
B vitamins (d -15 vs d 0 and d 5)		.18	.001	.006	.001	.001
BHV-1 (d 0 vs d 5)		.13	.001	.001	.001	.001
BHV-1 × B vitamin interaction		.35	.001	.003	.001	.001

^an = 6.

(d -15), although pyridoxal phosphate (**PLP**) tended to be higher in control calves. This may be due to variable hydrolysis of PLP to pyridoxal during prolonged storage before analysis (11 mo). Data are not available concerning B₆ vitamer stability in bovine plasma. Freezing rat plasma at -20°C for 55 wk did not affect B₆ content; however, these workers hydrolyzed the phosphate esters of the B₆ vitamers using a potato acid phosphatase before chromatography, so hydrolysis of PLP to pyridoxal was not assessed (Hefferan et al., 1986).

Coburn et al. (1984) previously measured much higher plasma PLP concentrations in three calves (402 ± 131 nM) than were detected in our calves using a very similar HPLC procedure. Pyridoxal phosphate was the primary B₆ vitamer in plasma, whereas pyridoxal was the primary vitamer in the current study. Nine samples from transport-stressed calves were analyzed simultaneously with samples from the current study. Samples from the stressed calves contained 126 ± 22 nM vitamin B₆, primarily in the form of PLP (111 ± 18 nM), whereas pyridoxal, pyridoxine, and the vitamin B₆ excretory product, 4-pyridoxic acid, averaged 8 ± 5 nM, 7 ± 7 nM, and 34 ± 2 nM, respectively (Dubeski, 1992). Compared to the transport-stressed calves, all samples from the current study were higher in total B₆ but lower in PLP and higher in pyridoxal. The samples from the transport-stressed calves were stored for only 6 mo. These data could be interpreted to suggest that longer storage causes hydrolysis of PLP to pyridoxal without affecting total B₆.

The effect of B vitamin injection was analyzed by contrasting values from d -15 vs d 0 plus d 5. B

vitamin treatment increased pyridoxal, 4-pyridoxic acid (**PA**), total B₆ ($P < .001$), and pyridoxine ($P < .006$). The interaction between BHV-1 infection and B vitamin treatment also was significant for these vitamers. Consequently, effects of B vitamin treatment will be discussed separately for d 0 and d 5.

On d 0, pyridoxal, pyridoxine, 4-pyridoxic acid, and total B₆ all were higher ($P < .002$) in plasma from +Vit than in plasma from control calves. Pyridoxal phosphate was not different ($P = .94$). Plasma from injected calves contained high concentrations of pyridoxine because pyridoxine hydrochloride was the B₆ source injected. The pyridoxine hydrochloride is converted gradually to the other B₆ vitamers. Plasma from +Vit calves also contained extremely high amounts of 4-pyridoxic acid, a major B₆ metabolite that in many species is excreted in the urine.

Compared to d -15, total B₆ on d 0 had decreased in control calves but increased in +Vit calves (interaction $P < .001$). Total B₆ may have fallen in controls because of a diminished supply with restricted feeding because duodenal B₆ supply is correlated with intake (Zinn et al., 1987). Conversely, +Vit calves received injections equal to eight times the estimated requirements. Because these estimates were designed to meet elevated dietary requirements associated with stress and disease, they probably exceeded requirements for the relatively unstressed calves in this study.

Plasma and tissue concentrations of B₆ vitamers are highly sensitive to changes in supply in non-ruminants. Blood and tissue concentrations were reduced 10 to 90% in rats by feeding a B₆-deficient diet for only 2 wk (Sampson and O'Connor, 1989). In

contrast, depriving food from healthy male dogs for 40 h decreased plasma PLP by 15% and increased pyridoxal by 20%; baseline concentrations again were achieved by 48 h after refeeding (Barnard et al., 1986). Similar comparisons are not available for ruminants but continued ruminal digesta outflow would be expected to attenuate fluctuations in supply and in plasma concentrations.

Various studies indicate that plasma PLP is derived primarily from recent dietary vitamin B₆ intake, particularly pyridoxine, and little pyridoxal is recycled through the liver into PLP in plasma (Coburn et al., 1992). Similar PLP concentrations in the two treatment groups on d 0 and higher concentrations of pyridoxal in +Vit calves on d 0 (645 vs 93 nM, $P < .001$) could indicate that PLP in plasma was degraded to pyridoxal during sample storage.

Total B₆ and all vitamers were lower in both control and +Vit calves on d 5 than on d 0. The most striking observation was the almost complete absence of pyridoxine from plasma of +Vit calves; only one calf had a detectable content of pyridoxine (57 nM). Similarly, both 4-pyridoxic acid and total B₆ were extremely decreased by BHV-1 infection in +Vit calves compared with control calves. Pyridoxal was decreased in both treatment groups as well.

Changes in concentrations of B₆ vitamers in this study indicate that B₆ metabolism was markedly affected by infection. Similarly, plasma PLP was sensitive to infection in hospital patients and was highly related to survival of critically ill surgical patients (Keniston et al., 1990). Changes in plasma concentrations during disease may reflect alterations in vitamin B₆ metabolism and increased requirements (Reynolds and Leklem, 1985; Merrill and Henderson, 1987). Pyridoxal phosphate is an essential cofactor for

numerous reactions in DNA, RNA, and protein synthesis that are crucial for the cell transformation and proliferation during the immune response, and furthermore, infection induces many catabolic reactions that require PLP (Beisel, 1977). Vitamin B₆ status in this study was very sensitive to a challenge to the immune system such as vaccination with live attenuated BHV-1. Increased susceptibility of stressed calves to disease and the frequent failure of stressed calves to respond appropriately to vaccination might be related to vitamin B₆ status.

Ascorbate

Plasma ascorbate tended to be decreased during the trial compared with before weaning (Table 7). Generally, plasma ascorbate concentrations were higher in these calves before weaning than in stressed calves, feedlot steers, and dairy cows (25.0, 27.8, and 23.8 μM , respectively) in a previous survey (Dubeski, 1992).

Plasma ascorbate concentrations were similar for the two treatment groups on d -15, before B vitamin injections began. Plasma ascorbate tended to decline in control and vitamin-injected calves between d -15 and d -8. This may be related to the negative energy balance; however, plasma ascorbate concentration did not decrease in control calves during the 3 d of food deprivation.

In humans, the major excretory route for ascorbate is the urine. When plasma concentration exceeds about 79.5 μM , substantial amounts of ascorbate spill over into urine (Jaffe, 1984). Although plasma concentrations in humans can be maintained as high as 227 μM with frequent and high ascorbate intakes, plasma ascorbate normally ranges from 45.4 to 79.5

Table 7. Ascorbic acid concentration in plasma (μM)

Day of experiment	Treatment mean ^a		SEM	Prob.
	Control	+Vit		
-45 Before weaning	59.2	59.2	2.3	—
-15 After weaning	55.1	50.5	2.8	.26
-8 Feed deprivation began	39.7	39.7	1.7	.94
-5 Feed deprivation ended	39.7	46.5	2.3	.09
0 BHV-1 inoculation	42.0	47.7	5.7	.001
5 Peak BHV-1 infection	21.6	32.4	3.4	.07
14 Final vitamin injection	36.9	42.0	4.0	.31
16 After vitamin injection	NA ^b	30.1	—	—
18 After vitamin injection	NA ^b	29.0	—	—
Contrast or change				
B vitamins (d -8 vs d -15)	-11.4	-5.7	3.4	.39
Feed deprivation (d -5 vs d -8)	0.0	5.7	2.8	.11
Refeeding (d 0 vs d -5)	2.8	58.4	4.5	.001
BHV-1 (d 5 vs d 0)	-20.4	-71.5	5.1	.001
Recovery (d 14 vs d 5)	14.8	7.9	3.4	.14

^an = 6.

^bNot analyzed.

μM (Jaffe, 1984); concentrations below $17.0 \mu\text{M}$ indicate a deficiency (Sauberlich, 1984a). Control calves in the current study had plasma ascorbate concentrations below the range expected for humans receiving adequate intakes of ascorbate.

Plasma ascorbate in +Vit calves was not markedly increased by the injection of 1,000 mg of sodium ascorbate every 48 h. Even though by d -8 the +Vit calves had received two ascorbate injections, plasma concentrations remained the same as in control calves. Carried nonspecifically in blood, ascorbate is taken up by organs using specific transport mechanisms that accumulate ascorbate (Jaffe, 1984). Ascorbate concentrations several hundred times those in plasma are found in the pituitary, adrenal, and thymus; concentrations are 10 to 100 times higher in small intestinal mucosa, lymph glands, lung, liver, spleen, and white blood cells (Sauberlich, 1984a). Hence, repletion of ascorbate by tissues might explain a lag before injections increased plasma ascorbate.

At the end of the food deprivation period (d -5), plasma ascorbate tended ($P < .09$) to be higher in the +Vit calves than in control calves. By this time, the +Vit calves had received four injections.

On d 0, plasma ascorbate was higher ($P < .001$) in +Vit than in control calves for the first time during the study. In both groups of calves, plasma ascorbate was lower ($P < .001$) at the time of peak BHV-1 infection (d 5) than on d 0, although concentrations still tended to be higher in the +Vit calves ($P = .07$). Jagos et al. (1977) reported that with calves from 2 to 3 mo of age, plasma concentrations were 28.4 ± 10.2 , 10.2 ± 6.2 , and $17.0 \pm 7.9 \mu\text{M}$ for healthy calves, calves with acute bronchopneumonia, and calves 3 wk after acute bronchopneumonia, respectively. Leukocytes contain very high concentrations of ascorbate compared with most tissues and plasma, and leukocytes can actively take up ascorbate during viral infection. Viral infections such as the common cold rapidly deplete the ascorbate content of leukocytes (Hume and Weyers, 1973).

With recovery from BHV-1 infection by d 14, plasma ascorbate concentrations in plasma increased in both groups of calves ($P < .001$). However, concentrations remained lower in controls than at any time prior to BHV-1 infection and in +Vit calves compared with concentrations on d -5 and d 0, suggesting that requirements as reflected by plasma ascorbate concentration may not have been fully satisfied. Perhaps either ascorbate requirements continued to be elevated or repletion of tissues or leukocytes was continuing at the expense of plasma.

After cessation of ascorbate injections, plasma ascorbate in +Vit calves dropped to concentrations lower than previously observed in control calves. Perhaps catabolism of ascorbate remains elevated when macrodoses of ascorbate are terminated, but whether scurvy results from such a withdrawal

remains controversial (Hornig and Moser, 1981). Long-term administration of ascorbate at high concentrations should be used with caution.

Implications

A mild respiratory (BHV-1) infection in steer calves markedly decreased plasma concentrations of vitamin B₆, vitamin B₁₂, pantothenic acid, and ascorbate but not folic acid. Restricting feed intake and depriving food for 3 d decreased plasma concentrations of vitamin B₆ and pantothenate, and increased plasma B₁₂, but did not markedly affect blood plasma folic acid and ascorbate concentrations. Like nonruminants, ruminants may be depleted in certain water-soluble vitamins when stress or immune challenge increase physiological requirements, especially when ruminal production is limited, conditions that occur often during the shipping and marketing process. Depletion of B vitamin and ascorbate status during shipping and marketing may contribute to the enhanced susceptibility of cattle to infectious disease during the first few weeks after arrival at feedlots. Based on plasma concentrations, status for certain vitamins critical to the immune response in cattle is decreased by an immune challenge, making supplementation desirable to counteract stress and disease.

Literature Cited

- Allgood, V. E., F. E. Powell-Oliver, and J. A. Cidlowski. 1990. Vitamin B₆ influences glucocorticoid receptor-dependent gene expression. *J. Biol. Chem.* 265:12424.
- Barnard, H. C., W.J.H. Vermaak, and G. M. Potgieter. 1986. The effect of acute prolonged starvation on the concentration of vitamin B₆ aldehyde derivatives in whole blood. *Int. J. Vit. Nutr. Res.* 56:351.
- Beisel, W. R. 1977. Magnitude of the host nutritional responses to infection. *Am. J. Clin. Nutr.* 30:1236.
- Coburn, S. P., and J. D. Mahuren. 1983. A versatile cation-exchange procedure for measuring the seven major forms of vitamin B₆ in biological samples. *Anal. Biochem.* 129:310.
- Coburn, S. P., J. D. Mahuren, and T. R. Guilarte. 1984. Vitamin B-6 content of plasma of domestic animals determined by HPLC, enzymatic and radiometric microbiological methods. *J. Nutr.* 114:2269.
- Coburn, S. P., J. D. Mahuren, M. S. Kennedy, W. E. Schaltenbrand, and D. W. Townsend. 1992. Metabolism of [¹⁴C]- and [³²P]pyridoxal 5'-phosphate and [³H]pyridoxal administered intravenously to pigs and goats. *J. Nutr.* 122:393.
- Crookshank, H. R., M. H. Elissalde, R. G. White, D. C. Clanton, and H. E. Smalley. 1979. Effect of transportation and handling of calves upon blood serum composition. *J. Anim. Sci.* 48:430.
- d'Offay, J. M., and B. D. Rosenquist. 1988. Combined effects of fasting and diet on interferon production and virus replication in calves infected with a vaccine strain of infectious bovine rhinotracheitis virus. *Am. J. Vet. Res.* 49:1311.
- Dubeski, P. L. 1992. B vitamins for cattle: Availability, plasma levels, and immunity. Ph.D. Dissertation. Oklahoma State Univ., Stillwater.
- Dubeski, P. L., J. M. d'Offay, F. N. Owens, and D. R. Gill. 1996. Effects of B vitamin injection on bovine herpesvirus-1 infection and immunity in feed-restricted beef calves. *J. Anim. Sci.* 74: 1367

- Dvorak, M. 1984. Ascorbic acid, stress resistance and reproduction in swine. In: *Workshop on Ascorbic Acid in Domestic Animals Proceedings*. p 80. Royal Danish Agricultural Society, Copenhagen.
- Fidanza, A., C. Bruno, A. De Cicco, S. Floridi, and L. Martoneli. 1978. Influenza di alte dosi di pantoteno di sodio sulla produzione deicorticosteroidi. *Boll. Soc. Ital. Biol. Sper.* 54:2248.
- Girard, C. L., J. J. Matte, and G. L. Roy. 1989. Serum folates in young dairy heifers. *Br. J. Nutr.* 61:595.
- Golub, M. S., and Gershwin, M. E. 1985. Stress-induced immunomodulation: What is it, if it is? In: G. P. Moberg (Ed.) *Animal Stress*. American Physiological Society, Bethesda, MD.
- Goodman, A. D. 1960. Studies on the effect of omega-methyl-pantothenic acid on corticosterone secretion in the rat. *Endocrinology* 66:420.
- Griffin, D. 1983. Feedlot disease losses. *Bovine Pract.* 16:88.
- Hefferan, T. E., Chrisley, B. M., and Driskell, J. A. 1986. Quantitation of B₆ vitamers in rat plasma by high-performance liquid chromatography. *J. Chromatogr.* 374:155.
- Hornig, D. H., and U. Moser. 1981. The safety of vitamin C intake in man. In: J. N. Counsell and D. H. Hornig (Ed.) *Vitamin C*. pp. 225-248. Applied Science Publishers, London.
- Hume, R., and E. Weyers. 1973. Changes in leucocyte ascorbic acid during the common cold. *Scot. Med. J.* 18:3.
- Jaffe, G. M. 1984. Vitamin C. In: L. J. Machlin (Ed.) *Handbook of Vitamins: Nutritional, Biochemical, and Clinical Aspects*. p 199. Marcel Dekker, New York.
- Jagos, P., J. Bouda, and R. Dvorák. 1977. Hladiny kyseliny askorbove pri bronchopneumonii telat. *Vet. Med. (Prague)* 22(3): 133.
- Keniston, R., S. Enriquez, Sr., and I. Delgado. 1990. Prognostic value of undeproteinized plasma pyridoxal 5'-phosphate levels in health and disease. In: K. Dakshinamurti (Ed.) *Vitamin B₆*. Ann. N.Y. Acad. Sci. 585:496.
- Maksymowych, A. B., V. Daniel, and G. Litwack. 1990. Pyridoxal phosphate as a regulator of the glucocorticoid receptor. *Ann. N. Y. Acad. Sci.* 585:438.
- McCormick, D. B. 1986. Vitamins. In: N. W. Teitz (Ed.) *Textbook of Clinical Chemistry*. p 960. W. B. Saunders, Philadelphia, PA.
- Merrill, A. H., Jr., and J. M. Henderson. 1987. Diseases associated with defects in vitamin B₆ metabolism or utilization. *Annu. Rev. Nutr.* 7:137.
- Mueller, C. B., and E. J. Thomas. 1975. Nutritional needs of the normal adult. In: W. F. Ballinger, J. A. Collins, W. R. Drucker, S. J. Dudrick, and R. Zeppa (Ed.) *Manual of Surgical Nutrition*. p 42. W. B. Saunders, Philadelphia, PA.
- Nauss, K. M., and P. M. Newberne. 1981. Effects of dietary folate, vitamin B₁₂ and methionine/choline deficiency on immune function. *Adv. Exp. Med. Biol.* 135:63.
- NRC. 1988. *Nutrient Requirements of Swine (9th Ed.)*. National Academy Press, Washington, DC.
- Reynolds, R. D., and J. E. Leklem (Ed.) *Current Topics in Nutrition and Disease*, Vol. 13. *Vitamin B-6: Its Role in Health and Disease*. Liss, New York.
- Roth, J. A. 1985. Cortisol as a mediator of stress-associated immunosuppression in cattle. In: G. P. Moberg (Ed.) *Animal Stress*. p 225. American Physiological Society, Bethesda, MD.
- Sampson, D. A., and D. K. O'Connor. 1989. Response of B-6 vitamers in plasma, erythrocytes and tissues to vitamin B-6 depletion and repletion in the rat. *J. Nutr.* 119:1940.
- SAS. 1985. *SAS User's Guide: Statistics (Version 5 Ed.)*. SAS Inst. Inc., Cary, NC.
- Sauberlich, H. E. 1984a. Ascorbic acid. In: *Nutrition Reviews: Present Knowledge in Nutrition*. 5th Ed. Chap. 18. The Nutrition Foundation, Washington, DC.
- Sauberlich, H. E. 1984b. Newer laboratory methods for assessing nutriture of selected B-complex vitamins. *Annu. Rev. Nutr.* 4: 377.
- Sauberlich, H. E. 1990. Vitamin B₆, vitamin B₁₂ and folate. *Adv. Meat Res.* 6:461.
- Shorthose, W. R., and J. R. Wythes. 1988. Transport of sheep and cattle. 34th Int. Cong. Meat Sci. Technol. Proc. (Part A):122.
- Song, W. O., A. Smith, C. T. Witter, B. W. Wyse, and R. G. Hansen. 1990. Determination of plasma pantothenic acid by indirect enzyme linked immunosorbent assay. *Nutr. Res.* 10:439.
- Von Tungeln, D. L. 1986. The effects of stress on the immunology of the stocker calf. *Bovine Pract.* 18:109.
- Wyse, B. W., C. Wittwer, and R. G. Hansen. 1979. Radioimmunoassay for pantothenic acid in blood and other tissues. *Clin. Chem.* 25:108.
- Zinn, R. A., F. N. Owens, R. L. Stuart, J. R. Dunbar, and B. B. Norman. 1987. B-vitamin supplementation of diets for feedlot calves. *J. Anim. Sci.* 65:267.